

Notes

Fibrin-Stabilizing Factor Inhibitors. 5. Primary Amines Related to Monotosylcadaverine¹

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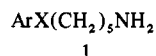
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Received November 15, 1971

A series of primary amines related to monotosylcadaverine has been prepared and tested as fibrin-stabilizing factor (FSF) inhibitors. The results indicate that an efficient specific inhibitor of FSF should have the general structure, $\text{ArX}(\text{CH}_2)_5\text{NH}_2$, where X should be a strongly electron-attracting group conjugated with the aromatic moiety.

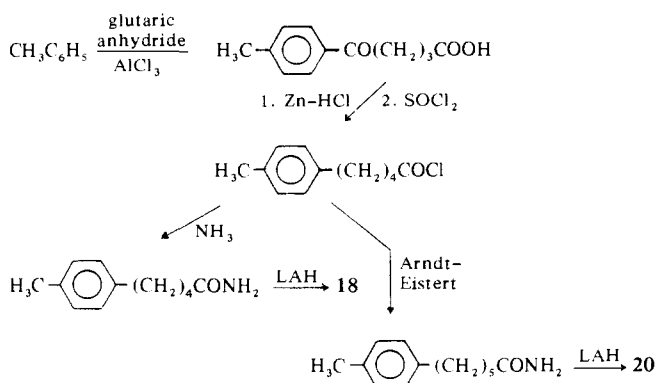
In previous papers of this series^{2a-c} we have published a number of new compounds which were tested as fibrin-stabilizing factor inhibitors (FSF inhibitors). From this work and from literature data^{3,4} we have concluded that a specific FSF inhibitor should have the general formula 1, where a pentyl chain, carrying a primary amino group at one end, is attached at its other end to an aromatic moiety (Ar) via a group X.



The primary amino group is essential for biologic activity^{2c,3} and the pentyl chain seems to be the optimal length for the intervening chain.^{2b,c,3} In this work we have investigated the influence of the group X on the activity.

Lorand, *et al.*,³ have discovered that monodansylcadaverine and monotosylcadaverine (2) were potent inhibitors of FSF. Substance 2 was chosen as a base for our investigations. A series of compounds was thus prepared where the SO_2NH moiety of 2 was replaced by other functional groups while the rest of the molecule was left intact.

Chemistry. The syntheses of the compounds were generally accomplished in a straightforward manner as described in the Experimental Section. Compounds 18 and 20 were obtained as indicated in Scheme I. The prepared compounds are presented in Table I.

Scheme I

Bioassays. The biological assays were carried out as previously described^{2a} and the inhibitory activity of each compound is expressed in relation to that of monodansylcadaverine (activity of monodansylcadaverine = 100).

Results and Discussion

The activities of the compounds are collected in Table I. These data show that the sulfonamides 2, 4, and 5 have the highest activity of the compounds tested. It is also apparent that among the compounds with other acyl groups, those with a carbonyl function directly conjugated with the aromatic ring have the best activity (*cf.* 7 and 9; 12 and 13). The compounds without the acyl group (15, 17, 18, 20) generally have low activity.

It is of interest to note that the combination of a sulfonamide with an aromatic moiety is necessary for high FSF-inhibitory activity. This was evidenced by the observation that the compound $\text{H}_2\text{NSO}_2(\text{CH}_2)_5\text{NH}_2$, which has no aromatic function, has an activity of less than 1% of that of monodansylcadaverine. As the SO_2NH group is strongly electron attracting, the aromatic nucleus of 2, 4, and 5 is expected to have an electron density that is low in comparison to that of the other test compounds. An electron-deficient aromatic nucleus in combination with the pentylamine as indicated in the general formula 1 thus appears to be a necessary structural requirement for high FSF-inhibitory activity.

The results of our studies obtained so far suggest that a specific FSF inhibitor should be attached to the enzyme near the active center via an electron-deficient aromatic nucleus. When the inhibitor is thus aligned in position, the side chain carrying the nucleophilic amino group must have a certain optimal length to permit the amino group to efficiently attack the carbonyl group of the thiol ester of the acyl-enzyme intermediate.⁴ This reaction will form an amide bond between a γ -carboxyl group of a Glu residue of fibrin and the NH_2 group of the inhibitor, thus inhibiting the fibrin cross-linking.

Experimental Section


General Comments. Melting points were determined with calibrated Anschütz thermometers in an electrically heated metal block. All crystal compounds were characterized by elemental analyses (C, H, N), which were within $\pm 0.4\%$ of the theoretical value, and infrared spectra, which were run for identification purposes on a Perkin-Elmer 237 spectrophotometer.

***N*-(5-Aminopentyl)-*p*-toluenesulfonamide (monotosylcadaverine) (2)** was prepared from TsCl and cadaverine by the procedure described for monodansylcadaverine;^{2c} yield 52%, mp for the hydrochloride 123–124° (from EtOH-Et₂O); lit.³ mp 123.5–124.5°.

***N*-(4-Cyanobutyl)-*N*-methyl-*p*-toluenesulfonamide (3).** TsCl (0.50 g, 2.6 mmoles) in CHCl_3 (45 ml) was added to a solution of 5-methylaminovalelonitrile⁵ (0.29 g, 2.6 mmoles) and Et_3N (1.0 g, 9.9 mmoles) in CHCl_3 (5 ml). The mixture was stirred at room temperature for 15 hr and was then washed with 5% aqueous NaHCO_3 (2 \times 50 ml) and with H_2O (2 \times 50 ml), dried (Na_2SO_4), and evaporated *in vacuo*. This yielded 0.65 g (94%) of a yellow oil which was used in the next step without further purification.

***N*-(5-Aminopentyl)-*N*-methyl-*p*-toluenesulfonamide (4).** Compound 3 (0.50 g, 1.9 mmoles) in EtOH (25 ml) and concentrated HCl (0.5 ml) was hydrogenated over PtO_2 at room temperature at an initial pressure of 3 kg/cm² until the theoretical amount of H_2 had been consumed. After filtration and evaporation of the filtrate, anhydrous Et_2O (100 ml) and EtOH

Table I. Physical Data and Biological Properties of the Compounds Studied

Compd No.	X	Yield, %	Mp, °C	H ₃ C-  -X(CH ₂) ₅ NH ₂		FSF-inhibitory activity ^a
				Formula		
2	SO ₂ NH	62	123-124	C ₁₂ H ₂₀ N ₂ O ₂ S·HCl	28	
4	SO ₂ NCH ₃	60	106-107	C ₁₃ H ₂₂ N ₂ O ₂ S·HCl	29	
5	NHSO ₂	34	147-149	C ₁₂ H ₂₀ N ₂ O ₂ S·HCl	20	
7	CONH	85	192-193	C ₁₃ H ₂₀ N ₂ O·HCl	5	
9	NHCO	74	180-182	C ₁₃ H ₂₀ N ₂ O·HCl	2	
11	NHCONH	83	182-183	C ₁₃ H ₂₁ N ₃ O·HCl	5	
12	CO ₂	18	118-120	C ₁₃ H ₁₉ NO ₂ ·HCl	8	
13	OCO	81	132-134	C ₁₃ H ₁₉ NO ₂ ·HCl	3	
15	NH	75	211-213	C ₁₂ H ₂₀ N ₂ ·2HCl	2	
17	NHCH ₂	70	214-215	C ₁₃ H ₂₂ N ₂ ·2HCl	2	
18		62	157-159	C ₁₂ H ₁₉ N·HCl	<1	
20	CH ₂	45	148-150	C ₁₃ H ₂₁ N·HCl	<1	

^aIn per cent of that of monodansylcadaverine, determined as previously described.^{2a}

(5 ml) were added. The pptd hydrochloride was collected and recryst from EtOH-Et₂O; yield 0.35 g (60%), mp 106-107°. *Anal.* (C₁₃H₂₂N₂O₂S·HCl) C, H, N.

5-(*p*-Tolylsulfamoyl)pentylamine (5) was prepared as previously described.^{2b}

N-(4-Cyanobutyl)-*p*-methylbenzamide (6). To a soln of *p*-toluoyl chloride (0.47 g, 3.0 mmoles) in anhyd Et₂O (50 ml) was added 5-aminovaleronitrile⁵ (0.59 g, 6.0 mmoles). The mixt was stirred at room temp for 1 hr and the pptd product was filtered off and washed with cold H₂O; yield 0.60 g (92%), mp 66-67° (from EtOH-H₂O). *Anal.* (C₁₃H₁₆N₂O) C, H, N.

N-(5-Aminopentyl)-*p*-methylbenzamide (7). Compd 6 (0.50 g, 2.3 mmoles) was hydrogenated over PtO₂ as described for 4. This yielded 0.50 g (85%) of the hydrochloride, mp 192-193° (from Me₂CO-EtOH). *Anal.* (C₁₃H₂₀N₂O·HCl) C, H, N.

6-Benzyloxycarbonylamino-hexanoyl-*p*-toluidide (8) was prepd from *N*-benzyloxycarbonyl-6-amino-hexanoic acid⁶ and *p*-toluidine by the mixed anhydride method as previously described;^{2a} yield 72%, mp 117-118° (from toluene). *Anal.* (C₂₁H₂₆N₂O₃) C, H, N.

6-Aminohexanoyl-*p*-toluidide (9). Compd 8 was hydrogenolyzed over Pd/C as previously described,^{2a} affording a yellow oil which was converted to the hydrochloride; yield 74%, mp 180-182° (from EtOH-Et₂O). *Anal.* (C₁₃H₂₀N₂O·HCl) C, H, N.

N-(4-Cyanobutyl)-*N'*-*p*-tolylurea (10). To a soln of *p*-tolyl isocyanate (0.68 g, 5.1 mmoles) in anhyd Et₂O (50 ml) was slowly added 5-aminovaleronitrile⁵ (0.50 g, 5.1 mmoles). The mixt was stirred for 1 hr at room temp and the product collected; yield 0.90 g (76%), mp 136-138° (from toluene). *Anal.* (C₁₃H₁₇N₃O) C, H, N.

N-(5-Aminopentyl)-*N'*-*p*-tolylurea (11). Compd 10 (0.46 g, 2.0 mmoles) was hydrogenated over PtO₂ as described for 4, affording 0.45 g (83%) of the hydrochloride, mp 182-183° (from Me₂CO-EtOH). *Anal.* (C₁₃H₂₁N₃O·HCl) C, H, N.

5-Aminopentyl *p*-Toluic Acid Ester (12). 5-Aminopentanol (2.1 g, 20 mmoles) was added to a soln of *p*-toluoyl chloride (3.1 g, 20 mmoles) and AcOH (1.2 g, 20 mmoles) in dry C₆H₆ (100 ml) and the mixt refluxed overnight. After cooling, Et₂O (100 ml) was added and the pptd product filtered off, suspended in 50 ml of 1 *M* Na₂CO₃, and extracted with CHCl₃ (3 × 50 ml). The organic ext was dried (Na₂SO₄) and evapd *in vacuo*, affording an oil which was dissolved in EtOH-Et₂O and pptd as hydrochloride; yield 0.90 g (18%), mp 118-120° (from EtOH-Et₂O). *Anal.* (C₁₃H₁₉NO₂·HCl) C, H, N.

p-Tolyl 6-Aminohexanoic Acid Ester (13). A mixt of 6-amino-hexanoyl chloride·HCl⁷ (4.5 g, 2.4 mmoles) and *p*-cresol (2.6 g, 2.4 mmoles) in dry C₆H₆ (75 ml) was stirred overnight at room temp. Anhyd Et₂O (100 ml) was then added and the pptd product filtered off and washed with Et₂O; yield 5.0 g (81%), mp 132-134° (from C₆H₆). *Anal.* (C₁₃H₁₉NO₂·HCl) C, H, N.

N-(4-Cyanobutyl)-*p*-toluidine (14). A soln of 5-chlorovaleronitrile (11.7 g, 100 mmoles) and *p*-toluidine (21.4 g, 200 mmoles) in *p*-xylene (250 ml) was refluxed for 24 hr. After cooling, the filtered soln was washed with 5% aqueous NaHCO₃ (2 × 150 ml) and with H₂O (2 × 150 ml), dried (Na₂SO₄), and evapd *in vacuo*. The residue was distd *in vacuo*, yielding 12.0 g (64%) of product, bp 156-158° (0.5 mm), mp 53-55° (from ligroin-EtOH). *Anal.* (C₁₂H₁₄N₂) C, H, N.

N-(5-Aminopentyl)-*p*-toluidine (15). The nitrile 14 (2.0 g, 10.6 mmoles) was refluxed with LAH (0.42 g, 11 mmoles) in anhyd Et₂O (150 ml) for 6 hr and then stirred overnight at room temp.

Aqueous 5 *M* NaOH (30 ml) was then added, and the Et₂O layer sepd. The alk phase was extd with Et₂O (3 × 100 ml) and the entire Et₂O extracts were dried (Na₂SO₄) and evapd. The residue was distd *in vacuo*, yielding a yellow oil, bp 155-160° (1.5 mm); mp of the dihydrochloride 211-213° (from EtOH-Et₂O), yield 2.1 g (75%). *Anal.* (C₁₂H₂₀N₂·2HCl) C, H, N.

N-(5-Cyanopentyl)-*p*-toluidine (16) was prepd from 6-bromo-capronitrile (4.5 g, 25.6 mmoles) and *p*-toluidine (5.5 g, 51 mmoles) as described for 14; yield 3.1 g (60%), mp 82-83° (from ligroin-EtOH). *Anal.* (C₁₃H₁₈N₂) C, H, N.

N-(6-Amino-hexyl)-*p*-toluidine (17) was prepd as described for 15; yield 70%, mp of the dihydrochloride 214-215° (from EtOH-Et₂O). *Anal.* (C₁₃H₂₂N₂·2HCl) C, H, N.

5-*p*-Tolylpentylamine (18). 5-*p*-Tolylpentanamide⁸ (0.50 g, 5 mmoles) was reduced with LAH (0.19 g, 5 mmoles) in anhyd Et₂O (150 ml) for 20 hr. The mixt was worked up as usual yielding a yellow oil which was pptd as the hydrochloride; mp 157-159° (from Me₂CO), yield 0.35 g (62%). *Anal.* (C₁₂H₁₉N·HCl) C, H, N.

6-*p*-Tolylhexanamide (19). To an ice-cold soln of CH₃N₂ (1.25 g, 30 mmoles) in Et₂O (60 ml) was slowly added a soln of 5-(*p*-tolyl)-pentanoyl chloride⁹ (2.0 g, 9.5 mmoles) in Et₂O (150 ml) over a period of 10 min. The mixt was stirred at room temp overnight and the Et₂O was distd off. Dioxane (25 ml), concd NH₃ (15 ml), and 10% aqueous AgNO₃ (3 ml) were added to the residual yellow oil and the mixt was refluxed for 2 hr. After cooling, ice (100 g) was added and the pptd amide collected; yield 1.15 g (59%), mp 119.5-120.5° (lit.¹⁰ mp 118.5°).

6-(*p*-Tolyl)hexylamine (20). This compd was prepd analogously to 18; yield 45%, mp of the hydrochloride 148-150° (from Me₂CO). *Anal.* (C₁₃H₂₁N·HCl) C, H, N.

5-Aminopentanesulfonamide was prepd as previously described.¹¹

Acknowledgment. We wish to thank Professor Laszlo Lorand at Northwestern University for most valuable discussions.

References

- (1) R. Lundén, A. Hartkoorn, J. L. G. Nilsson, P. Stenberg, J. Bruner-Lorand, and L. Lorand, *J. Med. Chem.*, **15**, 98 (1971) (paper 4).
- (2) (a) J. L. G. Nilsson, P. Stenberg, O. Eriksson, and R. Lundén, *Acta Pharm. Suec.*, **7**, 441 (1970); (b) P. Stenberg, J. L. G. Nilsson, O. Eriksson, and R. Lundén, *ibid.*, **8**, 415 (1970); (c) J. L. G. Nilsson, P. Stenberg, Ch. Ljunggren, O. Eriksson, and R. Lundén, *ibid.*, **8**, 497 (1971).
- (3) L. Lorand, N. G. Rule, H. H. Ong, R. Furlanetto, A. Jacobsen, J. Downey, N. Öner, and J. Bruner-Lorand, *Biochemistry*, **7**, 1214 (1968).
- (4) L. Lorand, *Thromb. Diath. Haemorrh.*, *Suppl.*, **39**, 75 (1970).
- (5) E. I. Vasilieva and R. Kh. Freidlina, *Izv. Akad. Nauk. SSSR Ser. Khim.*, 263 (1966).
- (6) H. Zahn and D. Hildebrand, *Chem. Ber.*, **90**, 320 (1957).
- (7) M. Rothe, H. Brüning, and G. Eppert, *J. Prakt. Chem.*, **8**, 323 (1959).
- (8) C. Willgerodt and W. Hambrecht, *ibid.*, **81**, 74 (1910).
- (9) R. Huisgen and V. Vossius, *Monatsh. Chem.*, **88**, 517 (1957).
- (10) R. Legros and P. Cangiari, *C. R. Acad. Sci.*, **250**, 147 (1960).
- (11) W. Griffin and D. H. Hey, *J. Chem. Soc.*, 3334 (1952).